

Evaluation of Antimicrobial and Antioxidant Properties of Biosynthesized Silver Nanoparticles Using Peel Extracts of *Citrus reticulata*

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Abstract: The plant extracts of *Citrus reticulata* well known to possess various biological properties. Hence, the present study was designed to evaluate antimicrobial and antioxidant activities of biosynthesized silver nanoparticles from peel extract of *Citrus reticulata*. The multiple antibiotic-resistant bacterial isolates were isolated from clinical samples of local hospital in and around Bangalore. Antibacterial activity of the biosynthesized silver nanoparticles using peel extracts of *Citrus reticulata* fruit was carried out by disc diffusion method. DPPH free radical scavenging method of antioxidant assay was performed using spectrophotometric method. In our study, the results of antibacterial activity revealed that silver nanoparticles synthesized by peel extract of *Citrus reticulata* fruit has effective antibacterial activities on the test isolates viz. *Enterobacter cloacae*, *Escherichia coli*, *Klebsiella pneumonia*, *Proteus mirabilis* and *Pseudomonas aeruginosa* as evidenced by the diameter of their zone of inhibition, and antioxidant activity of biosynthesized silver nanoparticles revealed effective free radical scavenging by silver nanoparticles. In summary, biosynthesized silver nanoparticles can be used as effective antibacterial agents even against multidrug resistant pathogenic bacteria, and possess significant free radical scavenging properties. In conclusion, our preliminary findings encourage to use eco-friendly silver nanoparticles for various medical and electronic applications.

Keyword: *Citrus reticulata*, Silver nano particles, Anti-microbial, Antioxidant

1. INTRODUCTION

A significant part of biosynthesis of nanoparticles is the utilization of plant concentrate to the biosynthesis reactions. Amalgamation of semi circular silver nanoparticles utilized a decontaminated apiin compound, extracted from henna leaf at ambient conditions. Natural methodologies utilizing microorganisms and plants or plant extracts for metal nanoparticle synthesis have been recommended as important options in contrast to chemical methods [1]. Plants have various reducing agents, for example, polyphenols and flavonoids etc. . . and are responsible for the reductions of Ag⁺ particles. These poly phenols and flavonoids are utilized as antimicrobial and antioxidant agents by the plants to shield themselves from different pathological conditions. Citrus fruits and derivatives have been notable to effectively affect human wellbeing inferable from their high concentration of Vitamin C and bioactive mixes, for example, phenolic acid, flavonoid, limonoid, carotenoid and fiber [2], [3].

The essential oils of *Citrus* sp. have wide varieties of functional properties viz. attractive aroma, a repellent agent against insects and animals, and antioxidant activities. Literature study evidenced the antimicrobial properties of citrus oils [4]. In addition, the advantages of citrus oils not only limited to food industry, but citrus oil in its oil and vapour form possess profound antimicrobial activities against wide range microorganisms viz. Gram-positive and Gram-negative bacteria [5].

Moreover, there are a large number of studies on plant essential oils regarding their antimicrobial properties in order to develop a source of antimicrobial ingredients for the food industry [6-8]. Furthermore, literature study revealed that *Citrus reticulata* has been reported to possess anti-bacterial [9], anti-fungal [10], anti-diabetic [11], cardio-protective [12],

anti-cancer [13], anti-arthritis [14], anti-inflammatory [15], anti-oxidant [16], anti-tubercular, and anti-anxiety medicinal properties [17].

With the rise and increment of microbial creatures impervious to numerous anti-infection agents, and the proceeding with accentuation on human services costs, various researchers have attempted to grow new, successful antimicrobial reagents liberated from resistance and cost. Such issues and needs have prompted the resurgence in the utilization of silver-based germ-killers that might be connected to broad spectrum activity and far lower penchant to initiate microbial resistance than antibiotics [18]. The antibacterial impacts of silver salts have been seen since ancient and silver is at present used to control bacterial development in an assortment of uses, including dental work, catheters and burn wounds [19-21]. With this scenario, the present study was designed to evaluate the antimicrobial and antioxidant properties of biosynthesized silver nano particles using peel extracts of *Citrus reticulata*.

2. MATERIALS AND METHODS

2.1 Collection of Pathogens

The multiple antibiotic-resistant isolates viz. *Escherichia coli*, *Proteus mirabilis*, *Klebsiella pneumonia*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* were isolated from clinical samples of local hospital in and around Bangalore and confirmed by various microscopic evaluation like Gram's staining [22]. Motility, capsule and spore formation was confirmed as per the procedure prescribed by Collins and Lyne [23]. All the bacterial pathogens were further confirmed by suitable biochemical tests [24] and used for antimicrobial activity studies.

2.2 Determination of Antimicrobial Activity

Antibacterial activity of the biosynthesized silver nanoparticles using peel extracts of *Citrus reticulata* fruit was carried out by disc diffusion method [25]. Nutrient agar medium plates were prepared, sterilized and solidified. After solidification bacterial cultures were swabbed on these plates. The sterile discs were dipped in silver nanoparticles solution (100 $\mu\text{g/ml}$) and placed in the nutrient agar plate and kept for incubation at 37°C for 24 hours. Zones of inhibition for (peel extracts of *Citrus reticulata* fruit and silver nitrate solution), silver nanoparticles and silver nitrate were measured. The experiments were repeated thrice and mean values of zone diameter were presented.

2.3 Antioxidant Assay:

Antioxidant assay for silver nanoparticles was carried out by modified method of Choi et al [26]. Different concentrations (0.025, 0.050, 0.125, 0.250, 0.5 and 1 mg/ml) of silver nanoparticles were individually mixed with 0.5 ml of 1 mM DPPH and incubated in dark for 30 minutes. After incubation the absorbance of the samples was determined at 517 nm against methanol as a blank by using UV-Visible spectrophotometer (Shimadzu, Japan). DPPH methanol reagent without sample was used as control and Vit.C was used as standard. The percentage of inhibition was calculated according to the following formula.

$$\% \text{ of inhibition} = \frac{[\text{Absorbancecontrol} - \text{Absorbancetest}]}{\text{Absorbancecontrol}} \times 100$$

3. RESULTS

The results of antibacterial activity portrayed that biosynthesized silver nanoparticles by using peel extract of *Citrus reticulata* fruit has effective antibacterial activities on the test isolates as indicated by the diameter of their zone of inhibition. The zone of inhibition was 22 mm for *Klebsiella pneumonia*, 21 mm for *Staphylococcus aureus*, 20 mm for *Proteus mirabilis*, 18 mm for *Enterobacter cloacae* & *Pseudomonas aeruginosa*, and 17 mm for *Escherichia coli*. Whereas the test shows the silver nitrate solution has no effect against tested isolates (Table 1 and Figure 1). The results of antioxidant activity of biosynthesized silver nanoparticles using peel extracts of *Citrus reticulata* depicted effective free radical scavenging by silver nanoparticles, and the antioxidant activity of silver nanoparticles was increased with increased concentration. (Figure 2).

Table. 1. The inhibitory activity of the biosynthesized silver nanoparticles using peel extracts of *Citrus reticulata* fruit against the test bacteria as demonstrated by diameters of zone of inhibition.

Bacterial Pathogens	Zone of Inhibition (mm)		
	Silver Nitrate Solution	Peel extract of <i>Citrus reticulata</i>	Peel extract of <i>Citrus reticulata</i> / Silver nanoparticles
<i>Enterobacter cloacae</i>	0	14	18
<i>Escherichia coli</i>	0	11	17
<i>Klebsiella pneumonia</i>	0	13	22
<i>Proteus mirabilis</i>	0	15	20

<i>Pseudomonas aeruginosa</i>	0	13	18
<i>Staphylococcus aureus</i>	0	15	21

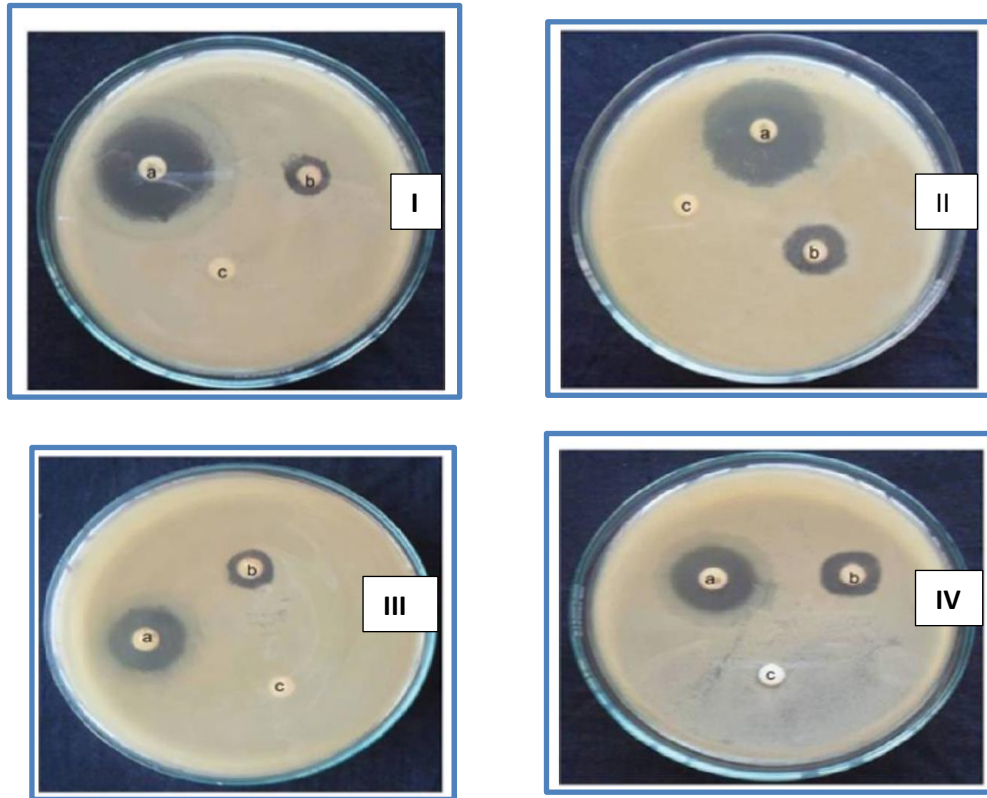


FIGURE 1: The antibacterial activity of biosynthesized silver nanoparticles from peel extracts of *Citrus reticulata* fruit against the test bacterium *Escherichia coli* [I], *Klebsiella pneumonia* [II], *Pseudomonas aeruginosa* [III], *Staphylococcus aureus* [IV].

Note: a-Citrus reticulata peel extract; b-silver nitrate solution; c-using test bacterium

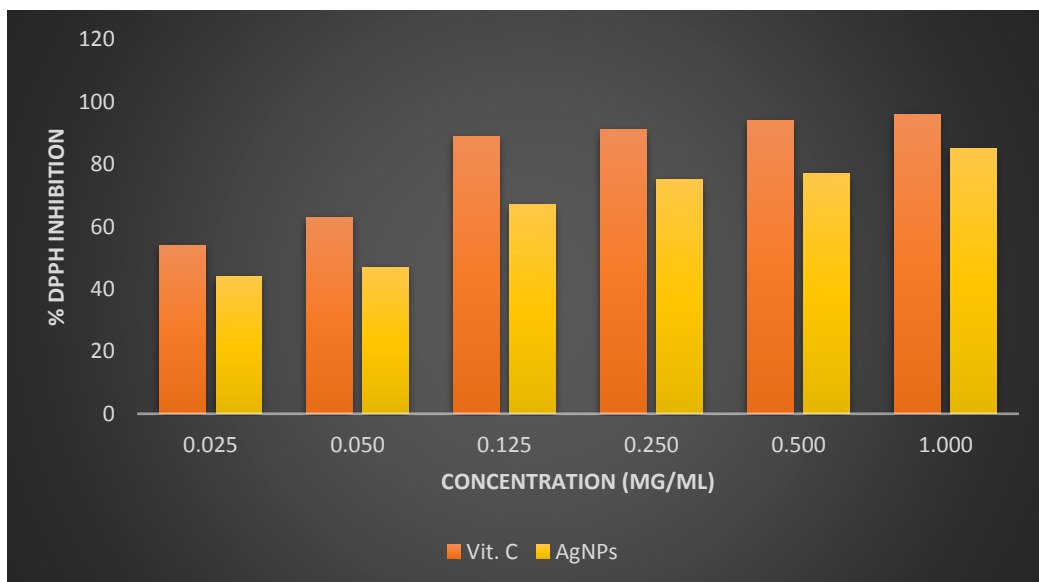


FIGURE 2: Antioxidant Activity of Biosynthesized Silver Nanoparticles

4. DISCUSSION

The results of antimicrobial activity delineated that biosynthesized silver nanoparticles utilizing peel extracts of *Citrus reticulata* fruit displayed viable antibacterial activities on the test isolates as demonstrated by the diameter of their zone of inhibition. Our outcomes showed that the biosynthesized silver nanoparticles using peel extracts of *Citrus reticulata* has another system to eliminate microorganisms not found in peel extracts of *Citrus reticulata* fruit alone. The results of the present investigation are in accordance with discoveries of Hindi et al [27].

The mechanism of the inhibitory effects of silver ions on microorganisms was partially known. previous studies demonstrated that the positive charge on the silver ions play a pivotal role for it's antimicrobial activity through the electrostatic attraction between negatively charged cell membrane of microorganism and positive charged nanoparticles [28-30]. In contrast, Sondi and Salopek-Sondi reported that the antimicrobial activity of silver nanoparticles on Gram-negative bacteria was dependent on the concentration of silver nanoparticle, and was closely associated with the formation of 'pits' in the cell wall of bacteria. Then, silver nanoparticles accumulated in the bacterial membrane caused the permeability, resulting in cell death. However, because those studies included both positively charged silver ions and negatively charged silver nanoparticles, it is insufficient to explain the antimicrobial mechanism of positively charged silver nanoparticles[31]. Amro et al suggested that metal depletion may cause the formation of irregularly shaped pits in the outer membrane and change membrane permeability, which is caused by progressive release of lipopolysaccharide molecules and membrane proteins [32]. Also, Sondi and Salopek-Sondi speculate that a similar mechanism may cause the degradation of the membrane structure of *E. coli* during treatment with silver nanoparticles [31]. Recently, Danilczuk et al reported silver-generated free radicals through the ESR study of silver nanoparticles that the antimicrobial mechanism of silver nanoparticles could be due to the formation of free radicals and subsequent free radical-induced membrane damage [33].

Antioxidant activity of biosynthesized silver nanoparticles from peel extracts of *Citrus reticulata* revealed effective free radical scavenging by silver nanoparticles. Our results are in accordance with previous findings[34-36]. Previous studies demonstrated that the antioxidant property could be due to development of reducing power. Reductones, which have strong reducing power, are generally believed not only to react directly with peroxides but also to prevent peroxide formation by reacting with certain precursors [37]. Silver nanoparticles are suggested to act as electron donors, reacting with free radicals to convert them to more stable products, which can terminate radical chain reaction. Furthermore, the reducing power of silver nanoparticles correlated well with the radical scavenging activity.

5. CONCLUSION

This study findings delineated that biosynthesized silver nanoparticles can be used as effective antibacterial agents even against multidrug resistant pathogenic bacteria, and possess significant free radical scavenging properties. Hence, our preliminary findings encourages to use eco-friendly silver nanoparticles for various medical and electronic applications.

REFERENCES

- [1] Kasthuri J, Veerapandian S, Rajendiran N. Biological synthesis of silver and gold nanoparticles using apiin as reducing agent. *Colloids and Surfaces B: Biointerfaces*. 2009;68(1):55-60.
- [2] Gorinstein S, Martín-Belloso O, Park YS, Haruenkit R, Lojek A, Číž M, Caspi A, Libman I, Trakhtenberg S. Comparison of some biochemical characteristics of different citrus fruits. *Food chemistry*. 2001;74(3):309-15.
- [3] Anagnostopoulou MA, Kefalas P, Kokkalou E, Assimopoulou AN, Papageorgiou VP. Analysis of antioxidant compounds in sweet orange peel by HPLC-diode array detection-electrospray ionization mass spectrometry. *Biomedical Chromatography*. 2005;19(2):138-48.
- [4] Subba MS, Soumithri TC, Rao RS. Antimicrobial action of citrus oils. *Journal of Food Science*. 1967;32(2):225-7.
- [5] Fisher K, Phillips C. Potential antimicrobial uses of essential oils in food: is citrus the answer?. *Trends in food science & technology*. 2008;19(3):156-64.
- [6] Mkaddem M, Bouajila J, Ennajar M, Lebrihi A, Mathieu F, Romdhane M. Chemical composition and antimicrobial and antioxidant activities of *Mentha (longifolia L. and viridis)* essential oils. *Journal of food science*. 2009;74(7):M358-63.
- [7] Zeng WC, Zhu RX, Jia LR, Gao H, Zheng Y, Sun Q. Chemical composition, antimicrobial and antioxidant activities of essential oil from *Gnaphalium affine*. *Food and Chemical Toxicology*. 2011;49(6):1322-8.
- [8] Salleh WM, Ahmad F, Yen KH, Sirat HM. Chemical compositions, antioxidant and antimicrobial activities of essential oils of *Piper caninum Blume*. *International journal of molecular sciences*. 2011;12(11):7720-31.

- [9] Viuda-Martos M, Ruiz-Navajas Y, Fernandez-Lopez J, Perez-Álvarez J. Antibacterial activity of lemon (*Citrus lemon* L.), mandarin (*Citrus reticulata* L.), grapefruit (*Citrus paradisi* L.) and orange (*Citrus sinensis* L.) essential oils. *Journal of food safety*. 2008;28(4):567-76.
- [10] Stange Jr RR, Midland SL, Eckert JW, Sims JJ. An antifungal compound produced by grapefruit and Valencia orange after wounding of the peel. *Journal of Natural Products*. 1993;56(9):1627-9.
- [11] Parmar HS, Kar A. Medicinal values of fruit peels from *Citrus sinensis*, *Punica granatum*, and *Musa paradisiaca* with respect to alterations in tissue lipid peroxidation and serum concentration of glucose, insulin, and thyroid hormones. *Journal of Medicinal Food*. 2008;11(2):376-81.
- [12] Kurowska EM, Manthey JA. Hypolipidemic effects and absorption of citrus polymethoxylated flavones in hamsters with diet-induced hypercholesterolemia. *Journal of agricultural and food chemistry*. 2004;52(10):2879-86.
- [13] Tanaka T, Makita H, Kawabata K, Mori H, Kakumoto M, Satoh K, Hara A, Sumida T, Tanaka T, Ogawa H. Chemoprevention of azoxymethane-induced rat colon carcinogenesis by the naturally occurring flavonoids, diosmin and hesperidin. *Carcinogenesis*. 1997;18(5):957-65.
- [14] Oben J, Enonchong E, Kothari S, Chambliss W, Garrison R, Dolnick D. Phellodendron and Citrus extracts benefit joint health in osteoarthritis patients: a pilot, double-blind, placebo-controlled study. *Nutrition journal*. 2009;8(1):1-9.
- [15] Emim JA, Oliveira AB, Lapa AJ. Pharmacological evaluation of the anti-inflammatory activity of a citrus bioflavonoid, hesperidin, and the isoflavonoids, dauricin and claussequinone, in rats and mice. *Journal of Pharmacy and Pharmacology*. 1994;46(2):118-22.
- [16] Tripoli E, La Guardia M, Giammanco S, Di Majo D, Giammanco M. Citrus flavonoids: Molecular structure, biological activity and nutritional properties: A review. *Food chemistry*. 2007;104(2):466-79.
- [17] Faturi CB, Leite JR, Alves PB, Canton AC, Teixeira-Silva F. Anxiolytic-like effect of sweet orange aroma in Wistar rats. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*. 2010;34(4):605-9.
- [18] Jones SA, Bowler PG, Walker M, Parsons D. Controlling wound bioburden with a novel silver-containing Hydrofiber® dressing. *Wound repair and regeneration*. 2004;12(3):288-94.
- [19] Silver S, Phung LT. Bacterial heavy metal resistance: new surprises. *Annual review of microbiology*. 1996;50(1):753-89.
- [20] Catauro MI, Raucci MG, De Gaetano F, Marotta A. Antibacterial and bioactive silver-containing Na₂O·CaO·2SiO₂ glass prepared by sol-gel method. *Journal of Materials Science: Materials in Medicine*. 2004;15(7):831-7.
- [21] Crabtree JH, Burchette RJ, Siddiqi RA, Huen IT, Hadnott LL, Fishman A. The efficacy of silver-ion implanted catheters in reducing peritoneal dialysis-related infections. *Peritoneal Dialysis International*. 2003;23(4):368-74.
- [22] Gram C. Ueber die isolirte Farbung der Schizomyceten in Schnitt-und Trockenpreparaten. *Fortschritte der Medicin*. 1884;2:185-9.
- [23] Collins CH. *Microbiological methods*. Microbiological methods.. 1967(2nd Edition).
- [24] Barrow GI, Feltham RK. *Manual for the identification of medical bacteria*. Cowan and steels: Cambridge, UK. 1993.
- [25] Cruickshank, R. (1968). *Medical Microbiology: A Guide to Diagnosis and Control of Infection*, Edinburgh/London: E. and S. Livingstone Ltd. 1968.
- [26] Choi CW, Kim SC, Hwang SS, Choi BK, Ahn HJ, Lee MY, Park SH, Kim SK. Antioxidant activity and free radical scavenging capacity between Korean medicinal plants and flavonoids by assay-guided comparison. *Plant science*. 2002;163(6):1161-8.
- [27] Hindi NK, Chabuck ZA, Hindi SK. Antibacterial Evaluation Of Aqueous Extract Of Four Citrus Species In Hilla, Iraq. *International Journal of Pharmacological Screening Methods*. 2014;4(1):45-6.
- [28] Hamouda T, Myc A, Donovan B, Shih AY, Reuter JD, Baker JR. A novel surfactant nanoemulsion with a unique non-irritant topical antimicrobial activity against bacteria, enveloped viruses and fungi. *Microbiological research*. 2001;156(1):1-7.
- [29] Dibrov P, Dzioba J, Gosink KK, Häse CC. Chemiosmotic mechanism of antimicrobial activity of Ag⁺ in *Vibrio cholerae*. *Antimicrobial agents and chemotherapy*. 2002;46(8):2668-70.
- [30] Dragieva I, Stoeva S, Stoimenov P, Pavlikianov E, Klabunde K. Complex formation in solutions for chemical synthesis of nanoscaled particles prepared by borohydride reduction process. *Nanostructured materials*. 1999;12(1-4):267-70.
- [31] Sondi I, Salopek-Sondi B. Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *Journal of colloid and interface science*. 2004;275(1):177-82.
- [32] Amro NA, Kotra LP, Wadu-Mesthrige K, Bulychev A, Mobashery S, Liu GY. High-resolution atomic force microscopy studies of the *Escherichia coli* outer membrane: structural basis for permeability. *Langmuir*. 2000;16(6):2789-96.
- [33] Danilczuk M, Lund A, Sadlo J, Yamada H, Michalik J. Conduction electron spin resonance of small silver particles. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2006;63(1):189-91.



- [34] Gao X, Zhang J, Zhang L. Hollow sphere selenium nanoparticles: their in-vitro anti hydroxyl radical effect. *Advanced Materials*. 2002;14(4):290-3.
- [35] Nie Z, Liu KJ, Zhong CJ, Wang LF, Yang Y, Tian Q, Liu Y. Enhanced radical scavenging activity by antioxidant-functionalized gold nanoparticles: a novel inspiration for development of new artificial antioxidants. *Free Radical Biology and Medicine*. 2007;43(9):1243-54.
- [36] Raghunandan D, Bedre MD, Basavaraja S, Sawle B, Manjunath SY, Venkataraman A. Rapid biosynthesis of irregular shaped gold nanoparticles from macerated aqueous extracellular dried clove buds (*Syzygium aromaticum*) solution. *Colloids and Surfaces B: Biointerfaces*. 2010;79(1):235-40.
- [37] Xia DZ, Yu XF, Zhu ZY, Zou ZD. Antioxidant and antibacterial activity of six edible wild plants (*Sonchus* spp.) in China. *Natural product research*. 2011;25(20):1893-901.