

Study on Isolation and Identification of Bacterium from Rhizosphere of Brinjal (*Solanum melongena* L.)

Jenifer Lolita C.¹, Keshamma E.², Shivashankrappa L. H³, Kavitha K. R.⁴

¹Associate Professor, Department of Botany, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

²Associate Professor, Department of Biochemistry, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

³Associate Professor, Department of Sericulture, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

⁴Associate Professor, Department of Botany & PG Studies, Nrupathunga University, Bengaluru, Karnataka, India

Abstract: Present study aimed to identify the bacteria isolated from rhizosphere of brinjal (*Solanum melongena* L.). The pure cultures of bacterial isolate from brinjal (*Solanum melongena* L.) were used to identify the bacteria. Identification of bacteria was done based on its morphological characteristics using reference strain viz. *Bacillus polymyxa* strain 10401 was obtained from France. Colony and morphological characters were observed under a light microscope, followed by Gram reaction and motility test. Results revealed that brinjal bacterial isolate had pearly colonies with smooth margin, constant in size, bead like and slightly raised from the surface of the culture plate. The BBI revealed rod shaped sporulating bacteria observed under light microscope. The BBI was Gram positive and turned yellow when L-Aniline 4-nitroanilide hydrochloride solution was added to the bacterial colony indicating the gram-positive nature of BBL. The bacteria were motile with the presence of single polar flagella as observed under a transmission electron microscope. The bacterial isolate which showed nitrogen fixing and phosphate solubilizing properties was identified as *Bacillus polymyxa* belonging to the class Eubacteriales, family Bacillaceae and genera *Bacillus*. In conclusion, for the first time the presence and identification of nitrogen fixing and phosphate solubilizing properties having bacteria was identified as *Bacillus polymyxa* on the rhizosphere of brinjal (*Solanum Melongena* L.).

Keyword: Morphological characteristics, *Solanum Melongena* L. *Bacillus polymyxa*, Brinjal Bacterial Isolate (BBI)

1. INTRODUCTION

In Karnataka, brinjal is cultivated in 16,602 hectares of land and has an average yield of 30-35 tons per hectare. Brinjal (*Solanum melongena* L.) is a member of the family Solanaceae and a native of India. It is an important vegetable crop of south India. The fruit is rich in vitamin A and vitamin C and is employed in Ayurveda system. The crop needs 369 kg of urea and 80 kg of phosphatic fertilizer per hectare. Although symbiotic nitrogen fixation especially legume-rhizobium system has been proved to be the best form of biological nitrogen fixation, associative nitrogen fixation cannot be ignored. Nitrogen fixation on the rhizoplane, phylloplane and stem have been attributed to the presence of diazotrophic bacteria associated with the roots, stem and leaves of plants [1].

There are many reports relating to the characterisation and identification of the nitrogen fixing bacteria associated with a wide variety of grasses and cereal crops. [2-8]. Hill et al., 1983 isolated and characterized rhizosphere bacteria of sweet potato [9]. Levanony et al. (1987) identified *Azospirillum brasiliense* in cereal roots using ELISA [10]. Rai and Guar, (1988) characterized *Azotobacter* associated with roots of wheat [8]. Lalande et al. (1989) identified rhizobacteria associated with maize as *Azospirillum* [10]. Lalande R (1989) identified *Azospirillum brasiliense* on surface and endosphere of wheat roots by immunogold labelling [10]. Brand et al. (1991) isolated a root colonizing rhizobacteria, which was characterized as *Pseudomonas* [11]. McInroy and Kloepper (1991) identified endophytic bacteria of maize and cotton [12].

Penot et al., 1991 [13], and McInroy and Kloepper, 1991 [12], characterized *Azospirillum* associated with maize cultivated in France, using biochemical tests. Agarwala et al. (1991) isolated associated bacteria from the interiors of many graminaceous plants, many of which were identified as *Azospirillum* [14]. Lukin (1990) observed spatial distribution of associated bacteria identified as *Azospirillum brasilense* the rhizosphere of barley plants [15]. Holguin et al. (1992) isolated and identified rhizobacteria associated with mangrove trees as *staphylococcus* [16]. Fages and Mulard (1986) identified the isolated bacteria from the rhizosphere of sunflower as *Azospirillum* [17].

With this viewpoints, in this study, efforts have been made to identify the brinjal bacterial isolate (BBI) from the rhizosphere of brinjal which reduce the addition of such high dose of nitrogen and phosphatic fertilizers, which promote growth and improve yield upon inoculation.

2. MATERIALS AND METHODS

2.1 Morphological characters

The isolated BBI was subcultured in solid and liquid nitrogen free liquid Burk's media. The pure cultures of BBI were used to characterize the bacteria. Identification of the dominantly associated bacterial isolate of brinjal (*Solanum melongena* L.) was done based on its morphological characteristics using a reference strain viz., *Bacillus polymyxa* strain 10401 obtained from France.

2.2 Colony characters and light microscopic studies

The bacterial identification required the observation of colony character and morphological characters under light microscopy. Gram reaction characteristics was assessed to identify the bacteria. The colony characters were recorded by observing 24 and 48 hr pure culture of the bacteria on solid Burk's media. The observations recorded were colony colour, margin and shape and the method followed was that of Subba Rao (1983) [18]. A drop of the 24 hr culture was smeared, stained and observed under oil immersion. Both vegetative and sporulating stages were observed.

2.3 Gram staining

Gram staining of the bacteria was done according to the procedure described in Bergey's 19-Manual of Determinative Bacteriology (1984). A drop of the pure culture was smeared on the slide, air-dried and fixed. The air-dried smear was treated sequentially with crystal violet, iodine, absolute alcohol and safranin. The gram reaction of the BBI was confirmed by adding 1% solution of L-aniline, 4-nitroanilide hydrochloride in tris buffer. When a drop of this solution was added on a colony of gram +ve bacteria, it turns yellow.

2.4 Motility test

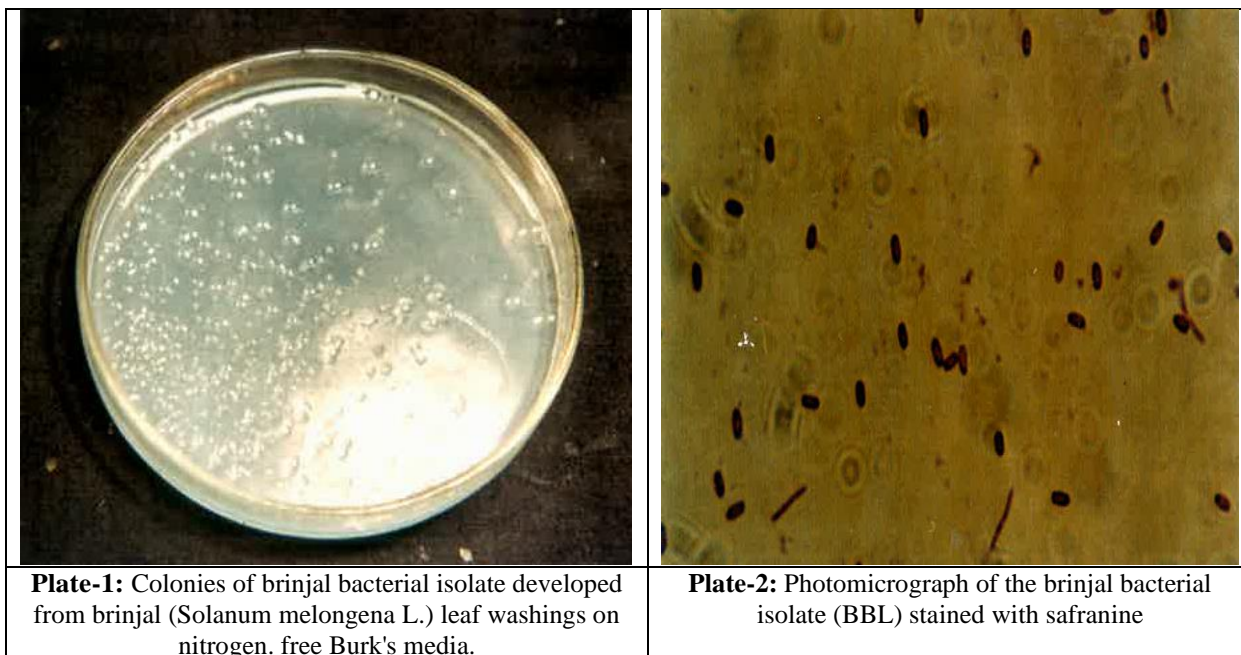
Stab cultures of the BBI in solid Burk's media were prepared and incubated at 37 °C. 24 hr and 48 hr cultures were examined for motility. Motility was confirmed by suspending a drop of 24hr culture in a cavity slide and observing under a light microscope [18].

3. RESULTS

3.1 Morphological characters

3.1.1 Colony characters and light microscopic studies

The BBI had pearly colonies, which were constant in size, bead like, and slightly raised from the surface of the culture plate (Plate-1). The colonies had smooth margin. Colonies of BBI showed good growth within 24 hours of inoculation on nitrogen free Burk's media. The light microscopic observations of the pure culture of the isolated associative BBI revealed it to be rod shaped sporulating bacteria (Plate-2).

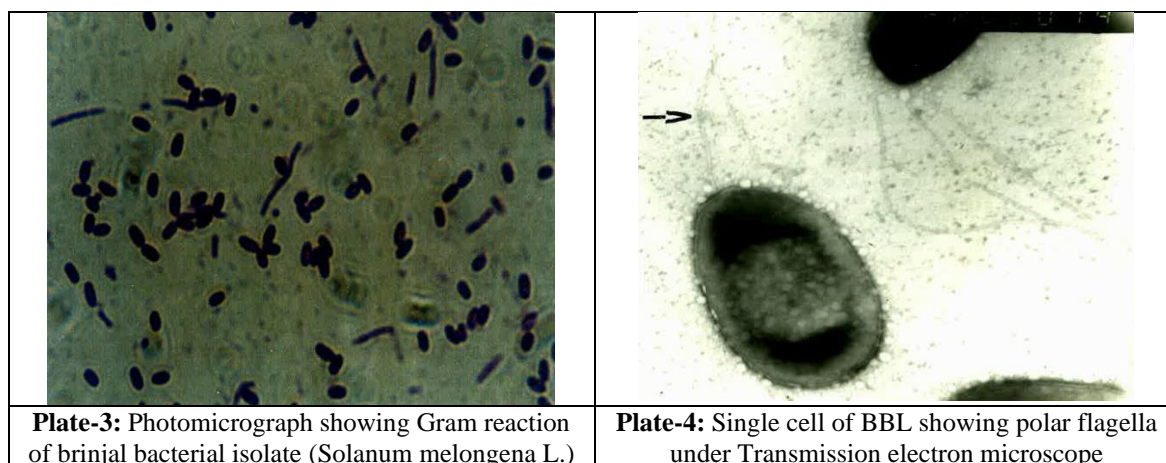


3.1.2 Gram staining

Gram staining of the bacteria revealed it to be gram positive which retained the crystal violet stain (Plate-3). L-Aniline 4-nitroanilide hydrochloride solution added to the bacterial colony turned yellow indicating the gram-positive nature of BBL.

3.1.3 Motility test

The spread of the bacteria in stab culture indicated the motility of the bacteria, observation by the hanging drop method and also confirmed the motility of BBL pure culture under transmission electron microscope (TEM) which revealed the presence of single polar flagella (Plate 4).



4. DISCUSSION

The bacterial isolate which showed nitrogen fixing and phosphate solubilizing properties was identified as *Bacillus polymyxa* belonging to the class Eubacteriales, family Bacillaceae and genera *Bacillus* (Bergey's Manual of determinative bacteriology 8th edition), which includes another phosphate solubilisers viz, *Bacillus macerans*. Identification was done based on its morphological characters. The colonies of *Bacillus polymyxa* were pearly and bead like in appearance, slightly raised from the surface of the culture plate and a had smooth margin. The isolate showed good growth within 24 hrs of inoculation. *Bacillus polymyxa* appeared to be rod shaped under light microscope. Growth promoting properties of *Bacillus polymyxa* have been reported by 20-Holl and Chanway (1997) in pine seedlings. These

studies further support the present finding of growth promotion by *Bacillus polymyxa* isolated from the roots of brinjal (*Solanum melongena* L.).

5. CONCLUSION

The study identified the bacterial isolate as *Bacillus polymyxa* which possess several plant growths promoting traits. This reveals the potential of this strain for biofertilizer application and commercial use as biocontrol agents in the field. Thus, this strain can perform close to its optimum potential. Future studies concerning commercial and field applications of integrated stable bio-formulations as effective biocontrol are needed.

REFERENCES

- [1] Dobereiner J. Forage grasses and grain crops. Methods for evaluating biological nitrogen fixation. 1980:535-6.
- [2] Dobereiner J, Marriel IE, Nery M. Ecological distribution of *Spirillum lipoferum* Beijerinck. Canadian Journal of Microbiology. 1976;22(10):1464-73.
- [3] Tarrand JJ, Krieg NR, Döbereiner J. A taxonomic study of the *Spirillum lipoferum* group, with descriptions of a new genus, *Azospirillum* gen. nov. and two species, *Azospirillum lipoferum* (Beijerinck) comb. nov. and *Azospirillum brasilense* sp. nov. Canadian journal of microbiology. 1978;24(8):967-80.
- [4] van Berkum P, Sloger C. Immediate acetylene reduction by excised grass roots not previously preincubated at low oxygen tensions. Plant Physiology. 1979;64(5):739-43.
- [5] Van Berkum P, Bohloul B. Evaluation of nitrogen fixation by bacteria in association with roots of tropical grasses. Microbiological reviews. 1980;44(3):491-517.
- [6] Silva DM, Ruschel AP, Matsui E, Lima Nogueira ND, Vose PB. Determination of the activity of N₂-fixing bacteria in sugarcane roots and bean nodules using tritiated acetylene reduction technique and electron microautoradiography. In Associative Nitrogen fixation Eds. 1981:145-152.
- [7] Levanony H, Bashan Y, Kahana ZE. Enzyme-linked immunosorbent assay for specific identification and enumeration of *Azospirillum brasilense* Cd. in cereal roots. Applied and Environmental Microbiology. 1987;53(2):358-64.
- [8] Rai SN, Gaur AC. Characterization of *Azotobacter* spp. and effect of *Azotobacter* and *Azospirillum* as inoculant on the yield and N-uptake of wheat crop. Plant and Soil. 1988;109(1):131-4.
- [9] Hill WA, Bacon-Hill P, Crossman SM, Stevens C. Characterization of N₂-fixing bacteria associated with sweet potato roots. Canadian journal of microbiology. 1983 ;29(8):860-2.
- [10] Lalonde, R., Bissonnette, N., Coutlee, D. and Antoun, H. (1989). Plant and Soil (Netherlands). 1989;117 (2): 207-218.
- [11] Brand I, Lugtenberg BJ, Glandorf DC, Bakker PA, Schippers B, De Weger LA. Isolation and characterization of a superior potato root-colonizing *Pseudomonas* strain. Bulletin OILB SROP (France). 1991.
- [12] McInroy JA, Klopper JW. Analysis of population densities and identification of endophyte bacteria of maize and cotton in the field. Bulletin OILB SROP (France). OILBSIRP. 1991:328- 331.
- [13] Penot I, Berges N, Guinguene C, Fages J. Characterization of *Azospirillum* associated with maize (*Zea mays*) in France, using biochemical tests and plasmid profiles. Canadian Journal of Microbiology. 1992;38(8):798-803.
- [14] Agarwala-Dutt R, Tilak KV, Rana JP. Isolation of *Azospirillum* from the interior of various parts of some graminaceous plants. Zentralblatt für Mikrobiologie. 1991;146(3):217-9.
- [15] Lukin S, Kozhevin P, Zviagintsev D. Spatial distribution of *Azospirillum brasilense* cells in the rhizosphere of barley plants. Mikrobiologičâ (Moskva, 1932). 1990(6):1090-3.
- [16] Holguin G, Guzman MA, Bashan Y. Two new nitrogen-fixing bacteria from the rhizosphere of mangrove trees: Their isolation, identification and in vitro interaction with rhizosphere *Staphylococcus* sp. FEMS Microbiology Letters. 1992;101(3):207-16.
- [17] Fages J, Mulard D. Isolation of rhizosphere bacteria and their effect on *Zea mays* in pots [*Azospirillum lipoferum*, *Enterobacter cloacae*, *Pseudomonas diminuta*]. Agronomie (France). Agronomie (France). 1988;1108(4):309-314.
- [18] Rao NS. Nitrogen-fixing bacteria associated with plantation and orchard plants. Canadian journal of microbiology. 1983;29(8):863-6.
- [19] Krieg NR, Holt JG. Bergey's manual of systematic bacteriology. Yi Hsien Publishing Co. 1984.
- [20] Holl FB, Chanway CP. Rhizosphere colonization and seedling growth promotion of lodgepole pine by *Bacillus polymyxa*. Canadian Journal of Microbiology. 1992;38(4):303-8.